Flow Cytometry
Director: Prof. Dr. Tobias Bopp

Instrumentation
- **Facs Symphony:** 33 parameter, 5 laser
- **Imagestream Mark II**
  - two orca flash cameras, 12 parameter 3 laser
  - three **FACS Aria** High speed Cell Sorters
  - up to 20 parameter and 5 laser
- **Moflo Legacy** in S2 cabinet coming up soon

Quantitative Proteomics
Director: Prof. Dr. Stefan Tenzer

Instrumentation
- **Two Waters nanoUPLC + Synapt G2-Si platforms**
  - High resolution (25,000 FWHM) and sensitivity
  - Integrated ion-mobility cell

Available Techniques & Methods
- Label-free quantitative proteome analysis
- Analysis of post-translational modifications
- Characterization of the MHC-ligandome
- Cell secretome analysis

Advanced Light Microscopy Unit
Director: Prof. Dr. Krishnaraj Rajalingam

Instrumentation
- **Leica SP8 CLSM, DMI8 TIRF and SP8 DIVE**

Available Techniques & Methods
- Confocal imaging: SP8 CLSM platforms (new HyD detectors) and widefield applications (DMI8 platforms)
- SP8 DIVE multiphoton confocal system → deep in vivo imaging
- DMI8 Infinity TIRF → high resolution imaging of dynamic processes (at the cell membrane)
- **In vivo** live cell imaging (DMI8 platforms equipped with environmental control box)
- offline LAS-X and Bitplane Imaris 9.1 software

Analysing, sorting and imaging in one centralized Core Facility

Analysis workstations:
- up to 32 cores; 128 gb ram
- FlowJo I-SNE
- Diva
- **ideas**

FZI Backbone drop in panels:
- T and B cell panel
- human and mouse
- ILC panel mouse
- Stem cell panel
- Human DC/Macrophages panel

FZI backbone panel feature:
- Streamlined panels with LID fluorescent dye
- Lineage dump Pe-Cy5 and LDA TAAO
- First color on laserline

Quantitative proteome profiling of human cancer cells using ion-mobility enhanced data-independent acquisition

Equipment of the Microscopy Unit

F-actin staining

1Kindly provided by K. Schütze and D. Nothmann

2Kindly provided by K. Biegadka and Dr. H. Yurugi